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(54) Title: CAROTENOID KETOLASE GENES AND GENE PRODUCTS, PRODUCTION OF KETOCAROTENOIDS AND METHODS OF MODIFYING CAROTENOIDS USING THE GENES (57) Abstract A purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 1 or 3, or has a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, as well as vectors and host cells containing them. Methods of use of the nucleic acid sequences to produce ketocarotenoid in host cells and methods of use of the nucleic acid sequences to modify the production of carotenoids in a host cell are included.			

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**CAROTENOID KETOLASE GENES AND GENE PRODUCTS,
PRODUCTION OF KETOCAROTENOIDS AND METHODS OF
MODIFYING CAROTENOIDS USING THE GENES**

BACKGROUND OF THE INVENTION

5 Carotenoids are widely distributed natural pigments that are responsible for many of the yellow, orange and red colors seen in living organisms. They have important commercial uses as coloring agents in the food industry, as feed and food additives, in cosmetics and as provitamin A precursors.

10 The plant species *Adonis aestivalis* produces flowers with petals that are deep red in color and nearly black at the base of the petals due to the accumulation of ketocarotenoid and other carotenoid pigments (Neamtu et al., *Rev. Roum. Biochim.* 6:157, 1969). This pattern of carotenoid accumulation accounts for the common name of some varieties of this species: summer pheasant's eye.

15 Among the carotenoids identified in the petals of the red petal varieties of these various species is the ketocarotenoid astaxanthin (3,3'-dihydroxy-4,4'-diketo-b,b-carotene; see Figure 1). Various other ketocarotenoids (see Figure 1) including 3-hydroxyechinenone (3-hydroxy-4-keto-b,b-carotene), adonirubin (3-hydroxy-4,4'-diketo-b,b-carotene) adonixanthin (3,3'-dihydroxy-4-keto-b,b-carotene) and isozeaxanthin (4,4'-dihydroxy-b,b-carotene; see T.W. Goodwin, *The Biochemistry of the Carotenoids*, 20 vol I. Plants, 2nd edition, 1980, page 147) have also been reported. The latter compound is consistent with speculation that the 4-hydroxy may be an intermediate in the formation of the 4-keto group.

SUMMARY OF THE INVENTION

25 There is appreciable interest in the biological production of carotenoids, in particular the orange-colored ketocarotenoids such as astaxanthin and canthaxanthin (Figure 1), and in the modification of carotenoid composition. For this reason, an *A. aestivalis* flower cDNA library was constructed and screened for cDNAs encoding enzymes (hereinafter referred to as "ketolases" although the specific biochemical activity has not yet been established) involved in the conversion of b-carotene into 30 orange compounds with absorption properties similar to those exhibited by common ketocarotenoids such as canthaxanthin (Figure 1). Two distinctly different *Adonis aestivalis* cDNAs were obtained from among a number of cDNAs that were selected on this basis.

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Thus, a first aspect of the present invention is a purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 1 or 3.

The invention also includes a purified nucleic acid sequence which encodes for 5 a protein having ketolase enzyme activity and having the amino acid sequence of SEQ ID NO: 2 or 4.

The invention also includes vectors which comprise any portion of the nucleic acid sequences listed above, and host cells transformed with such vectors.

Another aspect of the present invention is a method of producing a 10 ketocarotenoid in a host cell, the method comprising

inserting into the host cell a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and comprises (1) SEQ ID NO: 1 or 3 or (2) a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the heterologous nucleic acid sequence is operably linked 15 to a promoter; and

expressing the heterologous nucleic acid sequence, thereby producing the ketolase enzyme.

Another subject of the present invention is a method of modifying the production of carotenoids in a host cell, relative to an untransformed host cell, the method 20 comprising

inserting into a host cell which already produces carotenoids a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and comprises (1) SEQ ID NO: 1 or 3 or (2) a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the heterologous 25 nucleic acid sequence is operably linked to a promoter; and

expressing the heterologous nucleic acid sequence in the host cell to modify the production of the carotenoids in the host cell, relative to an untransformed host cell.

BRIEF DESCRIPTION OF THE DRAWINGS

30 A more complete appreciation of the invention and many of the attendant advantages thereof will be readily obtained as the same becomes better understood by

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reference to the following detailed description when considered in connection with the accompanying drawings.

Figure 1 illustrates structures and biochemical routes leading from b-carotene to various of the ketocarotenoids referred to in the text. Conversion of β -carotene to astaxanthin
5 by a hydroxylase enzyme (Hy) and a ketolase enzyme (keto) could proceed via any one or all of several possible routes depending on the order of the reactions.

Figure 2 illustrates the *beta* ring structure of b-carotene and various modifications of this parent ring that might be produced through the action of the products of the *A. aestivalis* ketolase cDNAs. Also shown is the structure of the *epsilon* ring, not found to be a
10 substrate for the *A. aestivalis* ketolases and present in carotenoids such as d-carotene, e-carotene, a-carotene and lutein.

Figure 3 illustrate results obtained with TLC (thin layer chromatography) separation of carotenoid pigments extracted from *E. coli* cultures, previously engineered to produce b-carotene, but that now also contain the *A. aestivalis* ketolase cDNAs and/or other
15 introduced genes and cDNAs. The Figure indicates the empty plasmid vector pBluescript SK- (SK-), the *Adonis aestivalis* ketolase 1 cDNA in this plasmid vector (Ad keto1), the *Haematococcus pluvialis* ketolase cDNA in this plasmid vector Hp keto), or the *Arabidopsis* β -carotene hydroxylase cDNA (At Ohase). Bands that were orange in color are shown here with a darker fill than those with a yellow color. Identities of
20 various bands are indicated to the right of the band.

Figure 4 illustrates the absorption spectrum of one of the orange carotenoids produced from b-carotene via the action of the *Adonis* ketolases and makes clear the similarity of the spectrum to that of canthaxanthin. Absorption spectra (in acetone) of β -carotene, canthaxanthin and an unknown orange product (orange band #1; the lower orange
25 band in the first lane of Figure 3) extracted from cultures after introduction of the *Adonis aestivalis* keto1 cDNA (SEQ ID NO: 1) in cells of *E. coli* that otherwise produce and accumulate β -carotene. The absorption spectrum of the unknown resembles that of canthaxanthin but the compound migrates to a position below echinenone on RP18

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TLC plates developed with a mobile phase of methanol:acetone (1:1 by volume). The absorption spectrum of orange band #2 also is similar to that of canthaxanthin but it migrates more rapidly than canthaxanthin indicating that it is probably a more polar compound.

5 Figure 5 shows SEQ ID NO: 5 (the sequence shown in this Figure includes SEQ ID NO: 1 and also includes some of the flanking DNA from the adaptor DNA and the multiple cloning site (MCS) of the library cloning vector, which sequences are shown in bold).

Figure 6 shows SEQ ID NO: 6 (the sequence shown in this Figure includes SEQ ID NO: 2 and also includes a translation of amino acids resulting from the adaptor DNA and
10 the multiple cloning site (MCS) of the library cloning vector and the start codon from the plasmid vector pTrcHis, which sequences are shown in bold and capitalized).

Figure 7 shows SEQ ID NO: 7 (the sequence shown in this Figure includes SEQ ID NO: 3 and also includes some of the flanking DNA from the adaptor DNA and the multiple cloning site (MCS) of the library cloning vector, which sequences are shown in bold).

15 Figure 8 shows SEQ ID NO: 8 (the sequence shown in this Figure includes SEQ ID NO: 4 and also includes a translation of amino acids resulting from the adaptor DNA and the multiple cloning site (MCS) of the library cloning vector and the start codon from the plasmid vector, which sequences are shown in bold and capitalized).

Figure 9 shows a "Gap" alignment of the two Adonis ketolase sequences of the
20 invention. A truncated version of SEQ ID NO: 1 is shown in this Figure for comparative purposes, and is designated SEQ ID NO: 9. The percentage identity was calculated to be 91.107.

Figure 10 shows a "Gap" alignment of SEQ ID NO: 2 and 4. The following results were found:

25 Gap weight: 12 average match: 2.912
 Length weight: 4 average mismatch: -2.003

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Quality: 1440 length: 307
Ratio: 4.691 gaps: 0
percent similarity: 92.182 percent identity: 90.228

Figure 11 shows a comparison between SEQ ID NO: 2 and the *Arabidopsis thaliana* β-
5 carotene hydroxylase enzyme (GenBank U58919) (SEQ ID NO: 10).

Figure 12A shows gDNA (SEQ ID NO: 11) immediately upstream of the cDNA of SEQ
ID NO: 3. The sequence was obtained from a PCR product generated using the
GenomeWalker kit of Clontech Laboratories, Inc. (1020 East Meadow Circle, Palo Alto,
CA 94303-4230) and nested primers specific to the ketolases of *Adonis aestivalis*
10 (cagaatcggtctgttctattagttcttcc (SEQ ID NO: 17) and caatttgaggaatatcaaggcccttgttctc
(SEQ ID NO: 18)). The termination codon upstream of and in-frame with initiation
codon (TAA at positions 204-206) is shown in bold. Initiation codon (ATG) is also
shown in bold.

Figure 12B (SEQ ID NO: 12) indicates that the full length polypeptide of SEQ ID NO:
15 4 begins with the amino acids MAA (shown in bold) immediately preceding the ketolase
sequence shown in Figure 8. A similar MAA amino acid sequence immediately
preceding SEQ ID NO: 1 is also expected.

Figure 13 shows an alignment of SEQ ID NO: 2, SEQ ID NO: 12, an *Arabidopsis* β-
20 carotene hydroxylase enzyme (predicted product of GenBank U58919) (SEQ ID NO:
13), a putative second *Arabidopsis* hydroxylase predicted by genomic DNA sequence
(GenBank AB025606; the exon/intron junctions were chosen with reference to the
product of the *Arabidopsis* β-carotene hydroxylase cDNA u58919) (SEQ ID NO: 14),
and two *Capsicum annuum* β-carotene hydroxylases (predicted products of GenBank
25 Y09722 and Y09225) (SEQ ID NO: 15 and 16).

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is directed to a purified nucleic acid sequence which

encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 1 or 3.

The invention also includes a purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and having the amino acid sequence of SEQ 5 ID NO: 2 or 4.

Two different but closely-related nucleic acids have been isolated. The sequences of the longest example of each are presented herein. Sequencing which has subsequently been conducted of upstream genomic DNA indicates that SEQ ID NO: 3 lacks bases encoding the first three amino acids (MAA; see Figure 12). Likely, 10 this is also the case for SEQ ID NO: 1, but the upstream genomic sequences have not yet been obtained for this nucleic acid.

The two different Adonis ketolases denoted in SEQ ID NO: 1 and 3 are similar in sequence, sharing about 91% identity, as determined by the Gap program discussed below (see Figure 9). The predicted amino acid sequences of the enzymes denoted in 15 SEQ ID NO: 2 and 4 share about 92% similarity and about 90% identity, also as determined by the Gap program (see Figure 10).

Therefore, it is clear that certain modifications of SEQ ID NO: 1 or 3 or SEQ ID NO: 2 or 4 can take place without destroying the activity of the enzyme. Note also that certain truncated versions of the cDNAs of SEQ ID NO: 1 or 3 were found to be 20 functional (i.e., these cDNAs retained the property of causing the conversion of b-carotene to orange compounds). Also, the *Arabidopsis* β -carotene hydroxylase (GenBank U58919), aligned with the ketolase SEQ ID NO: 2 in Figure 11, retains catalytic function when truncated to yield a polypeptide that lacks the first 129 amino acids (Sun et al., 1996). From the alignment in Figure 11, therefore, this would suggest 25 that the two ketolases of the invention retain catalytic activity after truncation to remove bases encoding the first 132 amino acids.

Thus, the present invention is intended to include those ketolase nucleic acid and amino acid sequences in which substitutions, deletions, additions or other modifications have taken place, as compared to SEQ ID NO: 1 or 3 or SEQ ID NO: 2 30 or 4, without destroying the activity of the ketolase enzyme. Preferably, the substitutions, deletions, additions or other modifications take place at those positions which already show dissimilarity between the present sequences. For SEQ ID NO: 1,

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as shown in Figure 9, these positions are as follows: positions 7, 20, 23, 35, 53, 63, 65, 67, 76, 78, 85, 86, 91, 107, 109-111, 135, 140, 144, 146, 160, 168, 217, 219, 241, 249, 254, 256, 271, 291, 296, 349, 389, 400, 406, 431, 448, 449, 460, 471, 499, 530, 589, 619, 643, 653, 654, 667, 679, 709, 731, 742, 784, 787, 836, 871, 883, 896, 911, 919,
5 928, 930, 939, 943, 967, 969, 978, 979, 982, 988, 995, 1005, 1006, 1012-1014, 1017, 1019-1021, 1023, 1025, 1049, 1050, 1054, 1060-1068, 1070-1073, 1075, 1094, 1100, 1101, 1106, 1107, 1109 and 1111-1176. For SEQ ID NO: 3, as shown in Figure 9, these positions are as follows: positions 7, 20, 23, 35, 53, 63, 65, 67, 76, 78, 85, 86, 91, 107, 109-111, 135, 140, 144, 146, 160, 168, 217, 219, 241, 249, 254, 256, 271, 291,
10 296, 349, 389, 400, 406, 431, 448, 449, 460, 471, 499, 530, 589, 619, 643, 653, 654, 667, 679, 709, 731, 742, 784, 787, 836, 871, 883, 896, 911, 919, 928, 930, 939, 943, 966, 967, 970, 979, 980, 983, 989, 996, 1006, 1007, 1013-1015, 1018, 1020-1022, 1024, 1026, 1050, 1051, 1055, 1062-1065, 1067, 1086, 1092, 1093, 1098, 1099, 1101 and 1103-1112.
15 For SEQ ID NO: 2 and 4, as shown in Figure 10, the following amino acids can be substituted or deleted, or additions or other modifications can be made, without destroying the activity of the ketolase enzyme: positions 7, 8, 12, 18, 21, 22, 25, 26, 36, 37, 45, 47-49, 56, 73, 83, 85, 97, 99, 130, 144, 150, 157, 166, 218, 244, 279, 299 and 304. Therefore, the present invention also intends to cover amino acid sequences
20 where such changes have been made.

In each case, nucleic acid and amino acid sequence similarity and identity is measured using sequence analysis software, for example, the Sequence Analysis, Gap, or BestFit software packages of the Genetics Computer Group (University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, Wisconsin 53705), MEGAlign
25 (DNAStar, Inc., 1228 S. Park St., Madison, Wisconsin 53715), or MacVector (Oxford Molecular Group, 2105 S. Bascom Avenue, Suite 200, Campbell, California 95008). Such software uses algorithms to match similar sequences by assigning degrees of identity to various substitutions, deletions, and other modifications, and includes detailed instructions as to useful parameters, etc., such that those of routine skill in the
30 art can easily compare sequence similarities and identities. An example of a useful algorithm in this regard is the algorithm of Needleman and Wunsch, which is used in the Gap program discussed above. This program finds the alignment of two complete

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sequences that maximizes the number of matches and minimizes the number of gaps. Another useful algorithm is the algorithm of Smith and Waterman, which is used in the BestFit program discussed above. This program creates an optimal alignment of the best segment of similarity between two sequences. Optimal alignments are found by
5 inserting gaps to maximize the number of matches using the local homology algorithm of Smith and Waterman.

Conservative (i.e. similar) substitutions typically include substitutions within the following groups: glycine and alanine; valine, isoleucine and leucine; aspartic acid, glutamic acid, asparagine and glutamine; serine and threonine; lysine and arginine; and
10 phenylalanine and tyrosine. Substitutions may also be made on the basis of conserved hydrophobicity or hydrophilicity (see Kyte and Doolittle, *J. Mol. Biol.* 157: 105-132 (1982)), or on the basis of the ability to assume similar polypeptide secondary structure (see Chou and Fasman, *Adv. Enzymol.* 47: 45-148 (1978)).

If comparison is made between nucleotide sequences, preferably the length of
15 comparison sequences is at least 50 nucleotides, more preferably at least 60 nucleotides, at least 75 nucleotides or at least 100 nucleotides. It is most preferred if comparison is made between the nucleic acid sequences encoding the enzyme coding regions necessary for enzyme activity. If comparison is made between amino acid sequences, preferably the length of comparison is at least 20 amino acids, more
20 preferably at least 30 amino acids, at least 40 amino acids or at least 50 amino acids. It is most preferred if comparison is made between the amino acid sequences in the enzyme coding regions necessary for enzyme activity.

While the two different Adonis ketolase enzymes of the present invention are similar in sequence, previously-described bacterial (Misawa et al., 1995), cyanobacterial
25 (Fernandez-Gonzalez et al., 1997), and green algal (*Haematococcus pluvialis*; Lotan et al., 1995; Kajiwara et al., 1995) β -carotene ketolase enzymes bear little resemblance to the Adonis ketolases, although certain histidine motifs and features of the predicted secondary structure are common to the polypeptides predicted by both groups (Cunningham and Gantt, 1998).

30 The present invention also includes vectors containing the nucleic acids of the invention. Suitable vectors according to the present invention comprise a gene encoding a ketolase enzyme as described above, wherein the gene is operably linked

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to a suitable promoter. Suitable promoters for the vector can be constructed using techniques well known in the art (see, for example, Sambrook et al., Molecular Cloning A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989; Ausubel et al., Current Protocols in Molecular Biology, Greene Publishing and Wiley Interscience, New York, 1991). Suitable vectors for eukaryotic expression in plants are described in Fray et al., (1995; *Plant J.* 8:693-701) and Misawa et al, (1994; *Plant J.* 6:481-489). Suitable vectors for prokaryotic expression include pACYC184, pUC119, and pBR322 (available from New England BioLabs, Beverly, MA) and pTrcHis (Invitrogen) and pET28 (Novagen) and derivatives thereof. The vectors of the present invention can additionally contain regulatory elements such as promoters, repressors, selectable markers such as antibiotic resistance genes, etc., the construction of which is very well known in the art.

The genes encoding the ketolase enzymes as described above, when cloned into a suitable expression vector, can be used to overexpress these enzymes in a host cell expression system or to inhibit the expression of these enzymes. For example, a vector containing a gene of the invention may be used to increase the amount of ketocarotenoids in an organism and thereby alter the nutritional or commercial value or pharmacology of the organism. A vector containing a gene of the invention may also be used to modify the carotenoid production in an organism.

Therefore, the present invention includes a method of producing a ketocarotenoid in a host cell, the method comprising

inserting into the host cell a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and comprises (1) SEQ ID NO: 1 or 3 or (2) a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the heterologous nucleic acid sequence is operably linked to a promoter; and

expressing the heterologous nucleic acid sequence, thereby producing the ketocarotenoid.

The present invention also includes a method of modifying the production of carotenoids in a host cell, relative to an untransformed host cell, the method comprising

inserting into a host cell which already produces carotenoids a vector comprising a heterologous nucleic acid sequence which encodes for a protein having

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ketolase enzyme activity and comprises (1) SEQ ID NO: 1 or 3 or (2) a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the heterologous nucleic acid sequence is operably linked to a promoter; and

expressing the heterologous nucleic acid sequence in the host cell to

5 modify the production of the carotenoids in the host cell, relative to an untransformed host cell.

The term "modifying the production" means that the amount of carotenoids produced can be enhanced, reduced, or left the same, as compared to an untransformed host cell. In accordance with one embodiment of the present invention,

10 the make-up of the carotenoids (i.e., the type of carotenoids produced) is changed *vis a vis* each other, and this change in make-up may result in either a net gain, net loss, or no net change in the amount of carotenoids produced in the cell. In accordance with another embodiment of the present invention, the production or the biochemical activity of the carotenoids (or the enzymes which catalyze their formation) is enhanced by the

15 insertion of the ketolase enzyme-encoding nucleic acid. In yet another embodiment of the invention, the production or the biochemical activity of the carotenoids (or the enzymes which catalyze their formation) may be reduced or inhibited by a number of different approaches available to those skilled in the art, including but not limited to such methodologies or approaches as anti-sense (e.g., Gray et al. (1992), *Plant Mol.*

20 *Biol.* 19:69-87), ribozymes (e.g., Wegener et al (1994) *Mol. Gen. Genet.* 1994 Nov 15;245(4):465-470), co-suppression (e.g. Fray et al. (1993) *Plant Mol. Biol.* 22:589-602), targeted disruption of the gene (e.g., Schaefer et al. *Plant J.* 11:1195-1206, 1997), intracellular antibodies (e.g., see Rondon et al. (1997) *Annu. Rev. Microbiol.* 51:257-283) or whatever other approaches rely on the knowledge or

25 availability of the nucleic acid sequences of the invention, or the enzymes encoded thereby.

Host systems according to the present invention preferably comprise any organism which is capable of producing carotenoids, or which already produces carotenoids. Such organisms include plants, algae, certain bacteria, cyanobacteria and

30 other photosynthetic bacteria. Transformation of these hosts with vectors according to the present invention can be done using standard techniques. See, for example, Sambrook et al., Molecular Cloning A Laboratory Manual, Cold Spring Harbor

Laboratory, Cold Spring Harbor, NY, 1989; Ausubel et al., Current Protocols in Molecular Biology, Greene Publishing and Wiley Interscience, New York, 1991.

Alternatively, transgenic organisms can be constructed which include the nucleic acid sequences of the present invention. The incorporation of these sequences can 5 allow the controlling of carotenoid biosynthesis, content, or composition in the host cell. These transgenic systems can be constructed to incorporate sequences which allow for the overexpression of the various nucleic acid sequences of the present invention. Transgenic systems can also be constructed which allow for the underexpression of the 10 various nucleic acid sequences of the present invention. Such systems may contain anti-sense expression of the nucleic acid sequences of the present invention. Such anti-sense expression would result in the accumulation of the substrates of the enzyme 15 encoded by the sense strand.

Having generally described this invention, a further understanding can be obtained by reference to certain specific examples which are provided herein for 15 purposes of illustration only and are not intended to be limiting unless otherwise specified.

EXAMPLE 1

Isolation of plant cDNAs that convert b-carotene into compounds with ketocarotenoid-like spectra

20 A flower cDNA library from the plant *Adonis aestivalis* was introduced into a strain of *Escherichia coli* engineered to accumulate the yellow carotenoid pigment β -carotene (see Cunningham et al., *Plant Cell* 8:1613-26, 1996). This strain of *E. coli* normally forms yellow colonies when cultures are spread on a solid agar growth medium. Ketocarotenoids that are derived from b-carotene, such as echinenone and 25 canthaxanthin (Figure 1), are, in contrast, orange to orange-red in color. Colonies that were orange rather than yellow in color were visually selected, and the DNA sequences of the *Adonis aestivalis* cDNAs within the plasmid vectors contained in these colonies were ascertained. Two distinct cDNAs were obtained from analysis of cDNA inserts in plasmids obtained from approximately 10 selected colonies. The DNA sequences of 30 these two ketolase cDNAs are presented herein.

The products produced by the ketolases of the invention which have been

expressed in a β -carotene-accumulating strain of *Escherichia coli* have not yet been identified. As many as 5 or 6 different colored bands, in addition to the substrate β -carotene, may readily be discerned by C₁₈ TLC separation (see Figure 3). To provide appropriate standards to assist in identification, an *H. pluvialis* ketolase and an 5 *Arabidopsis* β -carotene hydroxylase were separately introduced into the β -carotene- accumulating *E. coli* to produce echinenone (3-keto- β,β -carotene) and canthaxanthin (3,3'-diketo- β,β -carotene) or β -cryptoxanthin (4-hydroxy- β,β -carotene) and zeaxanthin (4,4'-dihydroxy- β,β -carotene). None of the compounds formed in the presence of the 10 ketolases of the invention (no difference was observed in products formed in the presence of the two different nucleic acid sequences of the invention) both migrate in the TLC system and have the absorption spectrum expected for echinenone, canthaxanthin, β -cryptoxanthin, or zeaxanthin. Two of the colored TLC bands produced in the presence of the Adonis ketolase cDNAs are orange in color. Orange band #1 has an absorption spectrum similar to that of canthaxanthin (see Figure 4) but migrates 15 in a position that indicates a polarity intermediate to echinenone and β -carotene. Orange band #2 also has an absorption spectrum like that of canthaxanthin but migrates in a position that indicates a polarity intermediate to canthaxanthin and zeaxanthin (see Figure 3). The absorption spectra and TLC results suggest that the two orange products could be desaturated at the 3-4 positions of both rings (3,4,- 20 didehydro; see Figure 2). Orange band #1 (see Figure 3) might then be 3,4,3',4'-tetradehydro- β,β -carotene. To substantially affect the absorption spectrum of the substrate β -carotene, any modifications very likely involve a carbon that lies in conjugation with the conjugated chain of carbon-carbon double bonds that constitute the chromophore (Goodwin, 1980; The Biochemistry of the Carotenoids, volume I; 2nd 25 edition, Chapman and Hall). For the spectra obtained, only the carbons at the number 4 position of the two rings appear to be plausible locations for modification. The multitude and TLC migrations of the yellow and orange products produced from the symmetrical β -carotene, however, also indicates that the enzymes of the invention carry out more than a single type of reaction. The apparent homology of the ketolases of the 30 invention to the *Arabidopsis* β -carotene hydroxylase would suggest that compounds with a hydroxyl at the 3 and/or 4 positions of one or both rings are another possible outcome (see Figure 2). In fact, such compounds have been identified in Adonis (see

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above), and it has long been conjectured that a hydroxyl at position 4 is an intermediate in the formation of the 4-keto (e.g. crustaxanthin, a 3,3',4,4' tetrahydroxy carotenoid that might be a precursor for astaxanthin in the exoskeleton of the lobster). The histidine motifs and secondary structure in common to the hydroxylase and ketolase enzymes 5 are characteristics of a large group of di-iron oxygenases whose members also include examples of desaturases (J. Shanklin, 1998, *Ann. Rev. Plant Physiol. Plant Mol. Biol.*), therefore a 3-4 desaturation (and/or perhaps a 2-3 desaturation in one or more of the yellow compounds) would also seem a plausible outcome.

To summarize the results of this example for the Adonis ketolases of the 10 invention, a number of different carotenoids, including two with ketocarotenoid-like spectra, are produced from β -carotene via the action of the products of either of the two different nucleic acids of the invention. These orange compounds appear to be the major products. Truncation and fusion of the cDNAs to a stronger promoter in the 15 vector pTrcHis (Invitrogen) was detrimental to growth of *E. coli* but did result in improved yield of the most polar orange product (orange band #2 in Figure 3). Introduction of a cyanobacterial ferredoxin did not change the yield or relative amounts 20 of the various products. Without being bound by theory, it may be that the ketocarotenoids produced in flower petals of Adonis actually include the as yet unidentified orange compounds that are produced in *E. coli* using the nucleic acids of the invention.

EXAMPLE 2

Substrate specificity of the Adonis ketolases

Carotenoids with ϵ rings are common in plants. The ϵ ring differs from the β ring only in the position of the double bond within the ring (Figure 2). The ϵ ring is reported 25 to be a poor substrate for the *Arabidopsis* β -carotene hydroxylase (Sun et al., 1996). The Adonis ketolase cDNAs were introduced into strains of *E. coli* engineered (Cunningham et al., 1996) to accumulate carotenoids with one or two ϵ rings (δ -carotene and ϵ -carotene), or the acyclic carotenoid lycopene. TLC analysis of acetone extracts revealed that these carotenoids were not modified by the Adonis ketolases. 30 as indicated by a lack of any new products formed. Products produced in *E. coli* engineered to accumulate zeaxanthin (Sun et al., 1996) appeared to be the same as

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for β -carotene accumulating cultures indicating that a 3-OH is likely to be one of the functional groups introduced to the b ring by the Adonis ketolases. The more polar orange band produced from β -carotene through the action of the Adonis ketolases (e.g., orange band 2 in Figure 3), therefore, could very well be 3,3'-dihydroxy-3,4,3',4'-
5 tetradehydro-b,b-carotene.

The references cited in the application, along with the following references, are incorporated by reference:

Bouvier F, et al. (1998) Xanthophyll biosynthesis: molecular and functional characterization of carotenoid hydroxylases from pepper fruits (*Capsicum annuum L.*).
10 Biochim Biophys Acta. 1391:320-8

Breitenbach J, et al. (1996) Expression in *Escherichia coli* and properties of the carotene ketolase from *Haematococcus pluvialis*. FEMS Microbiol Lett. 140:241-6

Cunningham FX Jr, Gantt E (1998) Genes and enzymes of carotenoid biosynthesis in plants. Ann Rev Plant Physiol Plant Mol Biol 49: 557-583

15 Fernandez-Gonzalez B, et al. (1997) A new type of asymmetrically acting beta-carotene ketolase is required for the synthesis of echinenone in the cyanobacterium *Synechocystis* sp. PCC 6803. J Biol Chem. 272:9728-33

Fraser PD, et al. (1997) In vitro characterization of astaxanthin biosynthetic enzymes. J Biol Chem. 1997272:6128-35

20 Fraser PD, et al. (1998) Enzymic confirmation of reactions involved in routes to astaxanthin formation, elucidated using a direct substrate in vitro assay. Eur J Biochem. 252:229-36

Harker M, et al. (1997) Biosynthesis of ketocarotenoids in transgenic cyanobacteria expressing the algal gene for beta-C-4-oxygenase, crtO. FEBS Lett. 404:129-34

- 15 -

Kajiwara S, et al. (1995) Isolation and functional identification of a novel cDNA for astaxanthin biosynthesis from *Haematococcus pluvialis*, and astaxanthin synthesis in *Escherichia coli*. *Plant Mol Biol*. 29:343-52

5 Lotan T, et al. (1995) Cloning and expression in *Escherichia coli* of the gene encoding beta-C-4-oxygenase, that converts beta-carotene to the ketocarotenoid canthaxanthin in *Haematococcus pluvialis*. *FEBS Lett*. 364:125-8

10 Misawa N, et al. (1995) Canthaxanthin biosynthesis by the conversion of methylene to keto groups in a hydrocarbon beta-carotene by a single gene. *Biochem Biophys Res Commun*. 209:867-76

Misawa N, et al. (1995) Structure and functional analysis of a marine bacterial carotenoid biosynthesis gene cluster and astaxanthin biosynthetic pathway proposed at the gene level. *J Bacteriol*. 177:6575-84

15 Miura Y, et al. (1998) Production of the carotenoids lycopene, beta-carotene, and astaxanthin in the food yeast *Candida utilis*. *Appl Environ Microbiol*. 64:1226-9

Shanklin J, et al. (1997) Mossbauer studies of alkane omega-hydroxylase: evidence for a diiron cluster in an integral-membrane enzyme. *Proc Natl Acad Sci U S A*. 94:2981-6

Shanklin J, Cahoon EB (1998) Desaturation and related modifications of fatty acids. *Ann Rev Plant Physiol Plant Mol Biol* 49: 611-641

20 Wang CW, et al. Engineered isoprenoid pathway enhances astaxanthin production in *Escherichia coli*. *Biotechnol Bioeng*. 1999 Jan 20;62(2):235-41.

I claim:

1. A method of producing a ketocarotenoid in a host cell, the method comprising
inserting into the host cell a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the 5 nucleic acid sequence of SEQ ID NO: 1 or 3, wherein the heterologous nucleic acid sequence is operably linked to a promoter; and
expressing the heterologous nucleic acid sequence, thereby producing the ketocarotenoid.
2. The method of claim 1, wherein the host cell is selected from the group 10 consisting of a bacterial cell, an algal cell and a plant cell.
3. A method of producing a ketocarotenoid in a host cell, the method comprising
inserting into the host cell a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the 15 heterologous nucleic acid sequence is operably linked to a promoter; and
expressing the heterologous nucleic acid sequence, thereby producing the ketocarotenoid.
4. The method of claim 3, wherein the host cell is selected from the group consisting of a bacterial cell, an algal cell and a plant cell.
- 20 5. A method of modifying the production of carotenoids in a host cell, relative to an untransformed host cell, the method comprising
inserting into a host cell which already produces carotenoids a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 1 or 3,
25 wherein the heterologous nucleic acid sequence is operably linked to a promoter; and
expressing the heterologous nucleic acid sequence in the host cell to modify the production of the carotenoids in the host cell, relative to an untransformed

host cell.

6. The method of claim 5, wherein the host cell is selected from the group consisting of a bacterial cell, an algal cell and a plant cell.

7. A method of modifying the production of carotenoids in a host cell, relative to an
5 untransformed host cell, the method comprising

inserting into a host cell which already produces carotenoids a vector comprising a heterologous nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has a sequence which encodes the amino acid sequence of SEQ ID NO: 2 or 4, wherein the heterologous nucleic acid sequence is operably
10 linked to a promoter; and

expressing the heterologous nucleic acid sequence in the host cell to modify the production of the carotenoids in the host cell, relative to an untransformed host cell.

8. The method of claim 7, wherein the host cell is selected from the group
15 consisting of a bacterial cell, an algal cell and a plant cell.

9. A purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 1.

10. A purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has the nucleic acid sequence of SEQ ID NO: 3.

20 11. A purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has a sequence which encodes the amino acid sequence of SEQ ID NO: 2.

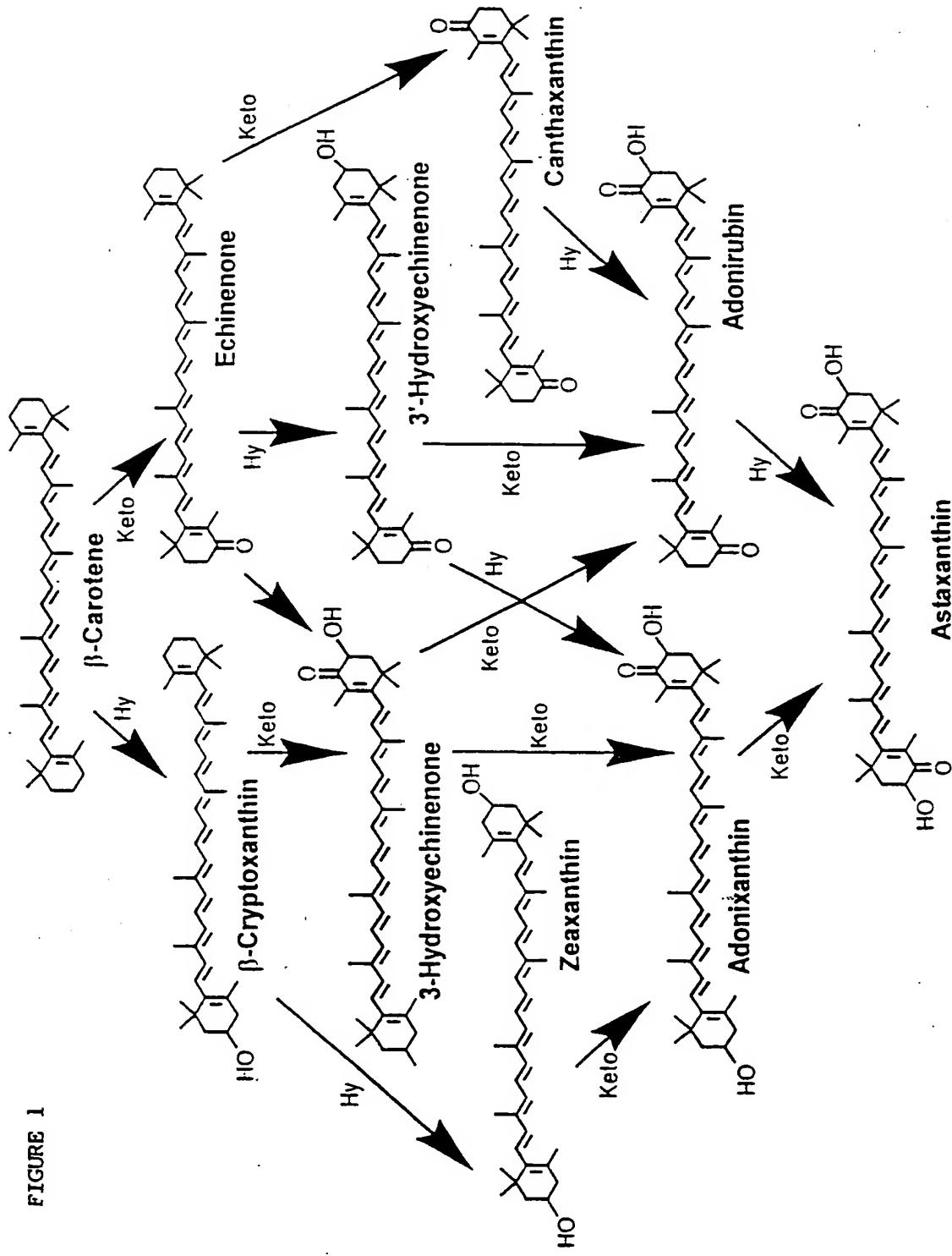
12. A purified nucleic acid sequence which encodes for a protein having ketolase enzyme activity and has a sequence which encodes the amino acid sequence of SEQ
25 ID NO: 4.

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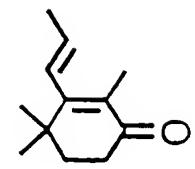
13. A vector which comprises the nucleic acid sequence of any one of claims 9-12, wherein the nucleic acid sequence is operably linked to a promoter.
14. A host cell which is transformed with the vector of claim 13.
15. The host cell of claim 14, wherein the host cell is selected from the group consisting of a bacterial cell, an algal cell and a plant cell.
16. The host cell of claim 14, wherein the host cell is a photosynthetic cell.
17. The host cell of claim 14, wherein the host cell contains a ketocarotenoid.
18. The host cell of claim 14, wherein the host cell contains modified levels of carotenoids, relative to an untransformed host cell.

10 19. A purified ketolase enzyme which is encoded by the amino acid sequence of SEQ ID NO: 2.

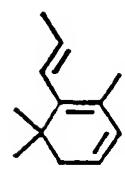
20. A purified ketolase enzyme which is encoded by the amino acid sequence of SEQ ID NO: 4.



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4-keto



3,4-didehydro

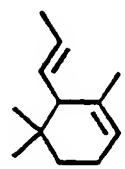
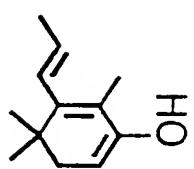
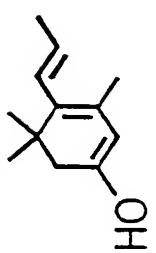
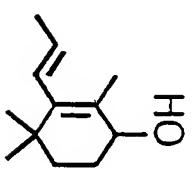
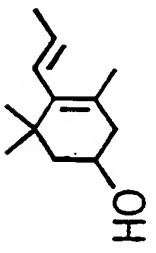
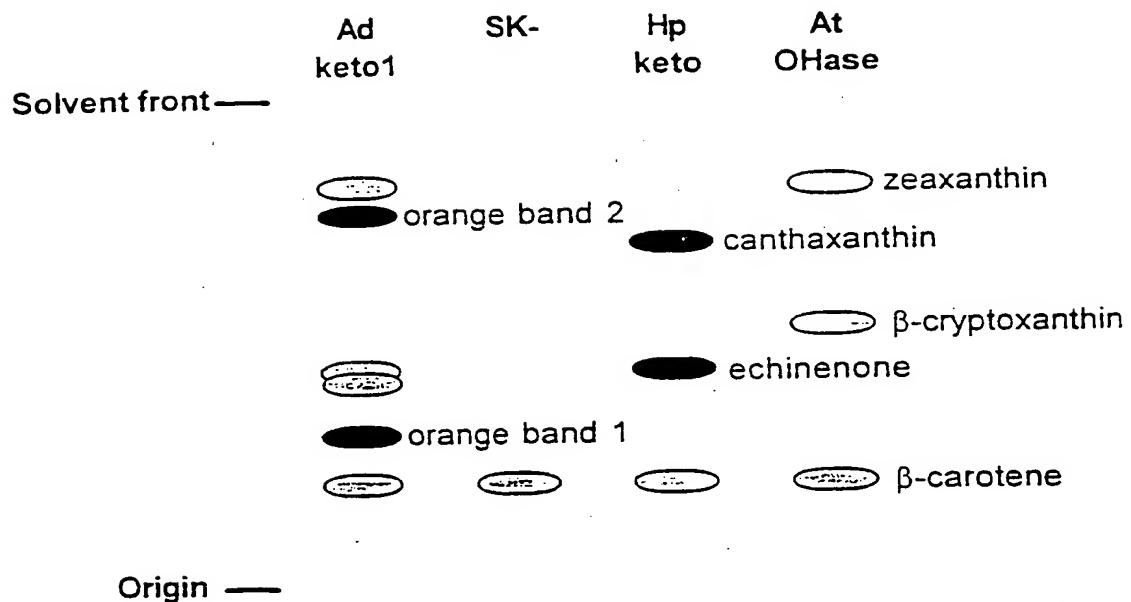
 ϵ ring β ring3,4-didehydro
4-hydroxy3,4-didehydro
3-hydroxy4-hydroxy
3,4-didehydro

FIGURE 2

FIGURE 3



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FIGURE 4

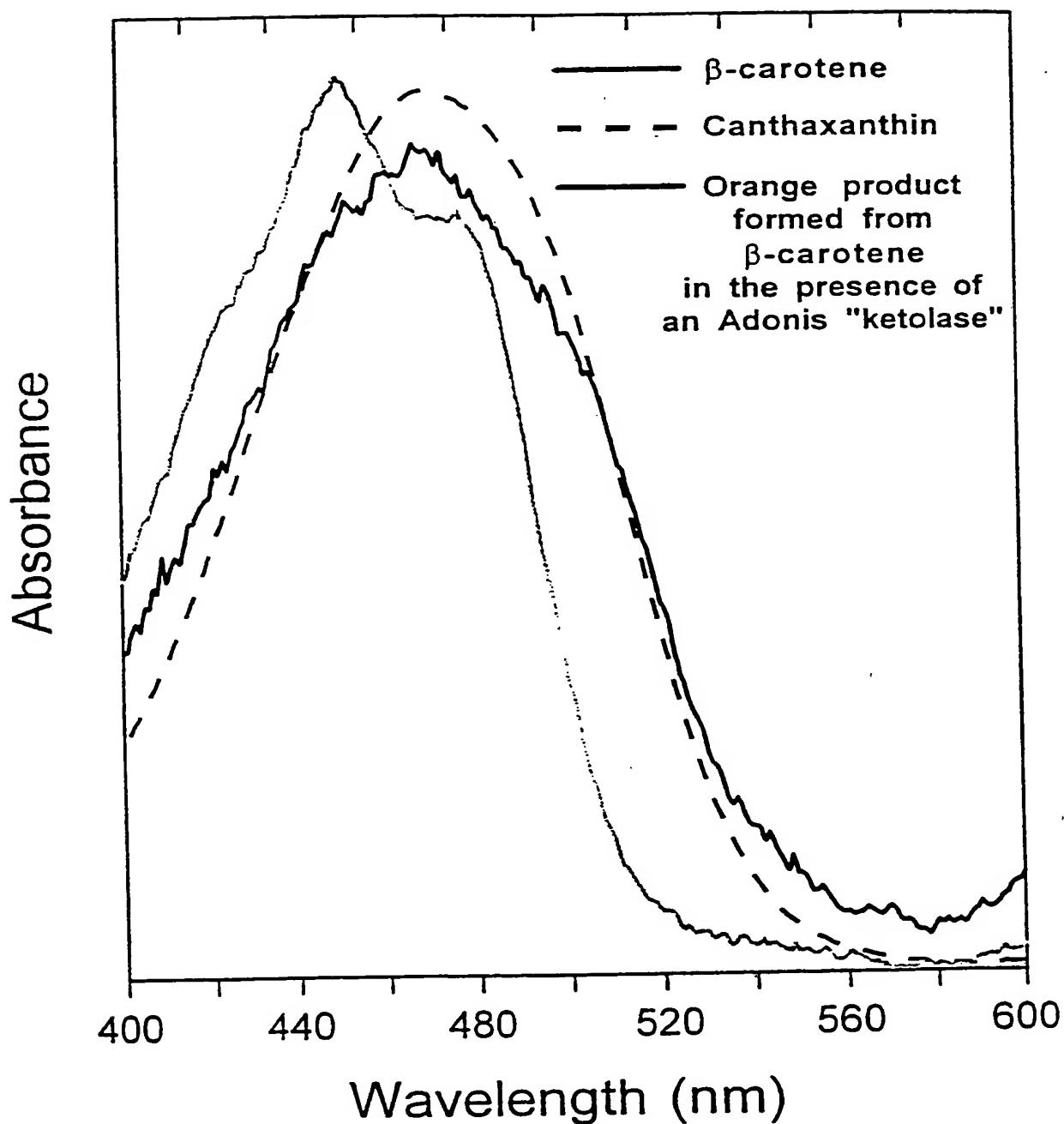


Figure 5 [SEQ ID NO: 5]

-23 ggg ctgcaggaat tcggcacgag
1 agcaatctca gtgttcagta caagttattc tttccacaag aatctcttgt
51 tgcactcaaa acaagacatt ctcAACCGCC catgtttgct cttctctcca
101 gttgtgggggg agtcgcctat gagaaagaaa aagacacatc gtgctgcatt
151 tatctgctct gttgcagaga gaacaaggaa ccttgatatt cctcaaattg
201 aagaagagga agagaacgag gaagaactaa tagaacagac ggattctggc
251 ataattccata taaagaaaaac gctagggggg aaacaatcaa gacggtccac
301 tggctccatt gtcgcaccccg tatcttgcct tgggatcctt tcaatgatcg
351 gacctgcgtt ttacttcaag tttcacggc taatggagtg tggagatatt
401 cctgtcgccag aaatggggat tacgtttgcc gcctttgttgc tgctgcgtat
451 tggcacggaa ttttgcgtt gatgggttca caaagaactc tggcacgatt
501 ctttggggta cattcacaag tctcaccata ggtcacgaaa aggccgcttc
551 gagttcaatg atgtgtttgc tattattaac gcgcttcctg ctattgctct
601 tatcaattat ggattctcaa atgaaggcct cttcccttggc gcctgctttg
651 gtaccggstt tggAACGACA gtctgtggca tggcttacat tttcttcac
701 aatggccccc cacaccgaag gttcccagta gggcttatttgc caaacgtccc
751 ttattccac aagctggctg cagctcacca aatccatcac tcaggaaaat
801 ttcagggstt accatttggc ctgttcccttgc gaccccagga attggaaagaa
851 gtaagaggag gcactgaaga attggagagg gtgatcagtc gtacagctaa
901 acgaacgcaa tcacatcatcatcatcatcatcatcatcatcatcatcatcatcat
951 agtttatcgg tgttacaagt cacacatggc tgtcgttgc gtaattcaaa
1001 gttaccatcac tcttttttag aatttttttt tgatgtatag gtcgcggagt
1051 tacggttaca aaggccaaat ctattgtgtt ggaattccat tattaaaaat
1101 aaaaattttaga gtttgcgtt ttatctggc atcaatatca atatatattat
1151 attaaagcaaa aaaaaaaaaaaa aaaaaaa ctcgag

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Figure 6 [SEQ ID NO: 6]

MGLQEFGTR

aisvfsts sys fhknlllhsk qdiln rpc11 fspvvvespm rkkk thraac
icsvaertrn ldipqieeee eneeelieqt dsgiihikkt lggkqsrrst
gsivapvscl gilmigpav yfkfsrlmec gdipvaemgi tfaafvaaai
gteflsgwvn kelwhd slwy ihkshh rsrk grfefndvfa iinalpaial
inygfsnegl lpgacf gtl gttvcgmayi flhnglshrr fpvglianvp
yfhklaaahq ihhsgkfqqv pfglflgpqe leevrggtee lervisrtak
rtqsst*

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Figure 7 [SEQ ID NO: 7]

-23 ggg ctgcaggaat tcggcacgag
1 agcaatttca gtgttcagtt caggttattc tttctacaag aatctcttgt
51 tggactcaaa accaaatatt ctcaaaccgg catgcctgct attctctcca
101 gttgtgatca tgtcgcctat gagaaagaaa aagaaacatg gtgatccatg
151 tatctgctcc gttgcagggga gaacaaggaa ctttgatatt cctcaaattg
201 aagaagagga agagaatgtg gaagaactaa tagaacagac cgattctgac
251 atagtgcata taaagaaaaac actagggggg aaacaatcaa aacggcccac
301 tggctccatt gtgcaccccg tatcttgtct tgggatcctt tcaatgattg
351 gacctgctgt ttacttcaag ttttacggc taatggaggg tggagatata
401 cctgttagcag aaatggggat tacgtttgcc acctttgttg ctgctgctgt
451 tggcacggag ttttgcag catgggttca caaagaactc tggcacgagt
501 ctttggta cattcacaag tctcaccatc ggtcacgaaa aggccgcttc
551 gagttcaatg atgtgtttgc tattattaac gcgcctcccg ctattgtct
601 tatcaattat ggattctcca atgaaggcct cttcccttggc gctgttttg
651 gtgtcggtct tggaaacaaca gtctgtggta tggcttacat ttttcttccac
701 aatggcctat cacaccgaag ttccccagta tggcttatttgc cgaacgtccc
751 ttatttccac aagctggctg cagctcacca aatacaccac tcaggaaaat
801 ttccagggtgt accatttggc ctgtttcccttgc gacccaagga atttggaaagaa
851 gtaagaggag gcactgaaga ttggagagg gtaatcagtc gtacaactaa
901 acgaacgcaa ccatctacct gaatcaattt ttttacatata ataaggttt
951 agtttatcgg ttttataaaaa tcacacatcc gtatcgaaaa agtaagtcaa
1001 agttaagata cttcccttctt agaatatttt ttgtatgtata ggtcgccggat
1051 atactgttac actattcggtt gtggaaattcc attataaaaaa aataaaaaaaa
1101 aaaaaaaaaaa aa ctcgag

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Figure 8 [SEQ ID NO: 8]

MGLQEFGTR

aisvfssgys fyknllldsk pnilkppc11 fspvvimspm rkkkkhgdpc
icsvagrtrn ldipqieeee enveelieqt dsdivhikkt lggkqskrpt
gsivapvscl gils migpav yfkfsrlmeg gdipvaemgi tfatfvaav
gteflsawvh kelweslwy ihkshhrsrk grfefndvfa iinalpaial
inygfsnegl lpgacfvgvgl gttvcgmayi flhnglshrr fpvwlianvp
yfhklaahq iihsgkfqgv pfglflgpke leevrggtee lervisrttk
rtqpst*

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Figure 9: Gap of SEQ ID NO: 9 and SEQ ID NO: 3

1 agcaatctcagtgttcagttacaagttattcttccacaagaatctcttgt 50
||||||| ||||||||||||| ||| ||||||||||| |||||||||||||
1 agcaatttcagtgttcagttcaggttattcttctacaagaatctcttgt 50

51 tgcactcaaaacaagacattctcaaccgcccattttgctctctcca 100
||| ||||||||| | | ||||||||| | ||||| | ||||| |||||
51 tggactcaaaaccaaatttctcaaaacccccatgcctgctattctctcca 100

101 gttgtggggggagtcgcctatgagaaagaaaaagacacatcggtgtgcattg 150
||||||| | | ||||||||| ||||||||| ||||| | ||| |
101 gttgtgatcatgtcgccatgagaaagaaaaagaaacatggtgatccatg 150

151 tatctgctttgtcagagagaacaaggaacccattgtatattcctcaaattg 200
||||||| | | ||||||||| ||||||||| ||||||||| ||||| |||||
151 tatctgctccgttgcagggagaacaaggaacccattgtatattcctcaaattg 200

201 aagaagaggaaagagaacgagggaaaactaataagaacacagacggattctggc 250
||||||| | | ||||||||| | | ||||||||| ||||| ||||| |
201 aagaagaggaaagagaatgtggaaagaactaataagaacacagacccattctgac 250

251 ataattcatataaaagaaaacgctaggggggaaacaatcaaacgggtccac 300
||| | | ||||||||| | | ||||||||| ||||| |||||
251 atagtgcataaaagaaaacactaggggggaaacaatcaaaacggccac 300

301 tggctccattgtcgacccgtatctgtcttggatccttcaatgatcg 350
||||||| | | ||||||||| ||||||||| ||||| ||||| |
301 tggctccattgtcgacccgtatctgtcttggatccttcaatgattg 350

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Figure 9 (cont.)

351 gacctgctgtttacttcaagtttacggctaattggagtgtggagatatt 400
||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
351 gacctgctgtttacttcaagtttacggctaattggagggtggagatata 400

401 cctgtcgccagaaatggggattacgttgcgccttggctgctgcgtat 450
||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |
401 cctgttagcagaaatggggattacgttgcaccccttggctgctgcgt 450

451 tggcacggaaattttgcaggatgggtcacaaaagaactctggcacgatt 500
||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |
451 tggcacggagttttgcagcatgggtcacaaaagaactctggcacgagt 500

501 ctttgtgg tacattcacaagtctcaccataggcacgaaaaggccgttc 550
||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
501 ctttgtgg tacattcacaagtctcaccatcggtcacgaaaaggccgttc 550

551 gagttcaatgatgtttgtattattaacgcgcgttccgtctattgtct 600
||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
551 gagttcaatgatgtttgtattattaacgcgcgttccgtctattgtct 600

601 tatcaattatggattctcaa atgaaggcctc ttccgttggagcgttgc 650
||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
601 tatcaattatggattctcca atgaaggcctc ttccgttggagcgttgc 650

651 gtaccggtcttggAACGACAGTCTGTGGCATGGCTTACATTTCTTCAC 700
||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
651 gtgtcggtcttggAACAGTCTGTGGATGGCTTACATTTCTTCAC 700

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Figure 9 (cont.)

701 aatggcctttcacaccgaagggtcccagtagggcttattgcaaacgtccc 750
||||||| ||||||| ||||| ||||| ||||| ||||| |||||
701 aatggccttatcacaccgaagggtcccagtaggttattgcgaacgtccc 750
. .
751 ttatttcacaagctggctgcagctcaccaaatccatcactcaggaaaat 800
||||||| ||||| ||||| ||||| ||||| ||||| |||||
751 ttatttcacaagctggctgcagctcaccaatacaccactcaggaaaat 800
. .
801 ttcagggtgtaccattggcctgttccttgacccaggaaattggaagaa 850
||||||| ||||| ||||| ||||| ||||| |||||
801 ttcagggtgtaccattggcctgttccttgacccaaggaaattggaagaa 850
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851 gtaagaggaggactgaagaattggagagggtgatcagtcgtacagctaa 900
||||||| ||||| ||||| ||||| ||||| |||||
851 gtaagaggaggactgaagagattggagaggtaatcagtcgtacaactaa 900
. .
901 acgaacgcaatcatctacaTGAatcaactctttacatttatgaggttt 950
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901 acgaacgcaaccatctaccTGAatcaatttttacatataaggttt 950
. .
951 agtttatcggtgtta.caagtcacacatttgtgtcggtgtägttaattcaa 999
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951 agtttatcggtgttataaaatcacacatccgtatcgtttagtaagtcaa 1000
. .
1000 agttaccataactcttttagaattttttgtatgtataggtcgccggag 1049
||||| ||||| ||| | | ||||| ||||| |||||
1001 agttaagataactcctttagaataattttgtatgtataggtcgccggat 1050

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Figure 9 (cont.)

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Figure 10: Gap of SEQ ID NO: 2 and SEQ ID NO: 4

1 AISVFSTSYSFHKNLLLHSKQDILNRPCLLFSPVVVESPMRKKKTHRAAC 50
||||| . ||| : ||||| ||| . ||| ||||| : ||||| ||| |
1 AISVFSSGYSFYKNLLLDSPNILKPPCLLFSPVVIMSPMRKKKHGDPC 50

51 ICSVAERTRNLDIPQIEEEEEENEEELIEQTDSGIHIKKTLGGKQSRRST 100
||||| ||||||| ||||| ||||| ||||| ||| : ||||| ||||| : |||
51 ICSVAGRTRNLDIPQIEEEEEENVEELIEQTDSDIVHIKKTLGGKQSKRPT 100

101 GSIVAPVSCLGILSMIGPAVYFKFSRLMECGDIPVAEMGITFAAFVAAAI 150
||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| ||||| :
101 GSIVAPVSCLGILSMIGPAVYFKFSRLMEGGDIPVAEMGITFATFVAAAV 150

151 GTEFLSGWVHKELWHDSLWYIHKSHHRSRKGRFEFNDVFAIINALPAIAL 200
||||| ||||| : ||||| ||||| ||||| ||||| ||||| ||||| |||||
151 GTEFLSAWVHKELWESLWYIHKSHHRSRKGRFEFNDVFAIINALPAIAL 200

201 INYGFSNEGLLPGACFGTGLGTTVCGMAYIFLHNGLSHRRFPVGLIANVP 250
||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||
201 INYGFSNEGLLPGACFGVGLGTTVCGMAYIFLHNGLSHRRFPVWLIANVP 250

251 YFHKLAAAHQIHSGKFQGVPGFLGPQELEEVRGGEELERVISRTAK 300
||||| ||||| ||||| ||||| ||||| . ||||| ||||| ||||| |||||
251 YFHKLAAAHQIHSGKFQGVPGFLGPKELEEVRGGEELERVISRTTK 300

301 RTQSST* 307
||| |||
301 RTQPST* 307

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Figure 11: Gap of SEQ ID NO: 2 and *Arabidopsis* β -carotene hydroxylase (SEQ ID NO: 10)

1 AISVFSTSYSFHKNLLLHSKQDILNRPCLLFSPVVVESPMRKKKTHRAAC 50
 . || . | || . | . | . : | .
1 MAAXLSTAVTFKP...LHRSFSSSSSTDFLRLLPKSLSGFSPSLRFKRF SV 47
 . . .
51 ICSVAERTRNLDIPQIEEEEENEELIEQTDSGIIHIKKTLGGKQSRRST 100
 | || . | | | | : : : . | . | || |
48 CYVVEERRQNSPIENDERPESTSSTNAIDAELYLALRLAEKLERKKSERST 97
 . .
101 GSIVAPVSCLGILSMIGPAVYFKFSRLMECGDIPVAEMGITFAAFVAAA 150
 | | . | || | | ||::|| | | | . || | || | || | ||:
98 YLIAAMLSSFGITSMAVMAYYRF SWQMEGGEISMLEMFGTFALSVGA AV 147
 . .
151 GTEFLSGWVHKELWHDSLWYIHKSHRSRKGRFEFNDVFAIINALPAIAL 200
 | || . | | : | || | | . | . || : | . | | | | | | | |
148 GMxEFWARWAHRALWHASLWNMHESHHKPREGPFELNDVFAIVNAGPAIGL 197
 . .
201 INYGFSNEGLLPGACFGTGLGTTVCGMAYIFLHNGLSHRRFPVGLIANVP 250
 : . || | | - | | | | | | | | | | | | | | | | | | | | | |
198 LSYGFFNKGLVPGLCFGAGLGITVFGIAYMFDVHDGLVHKRFPVGPIADVP 247
 . .
251 YFHKLAAAHQIHHSGKFQGVPGFLGPQELEEVVRGGTEELERVISRTAK 300
 | | . | | | | : | | . | | | | : | | | | | | | | | | | | |
248 YLRKVAAAHQLHHTDKFNGVPYGLFLGPKELEEV.GGNEELDKEISRRRIK 296
 . .
301 RTQSST*..... 307
 . .
297 SYKKASGSGSSSS* 311

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Figure 12A (SEQ ID NO: 11)

1 CATAACCATAAA ATAGTAGAGG ACAACCTACA AACCAACCAC CAGAAACCTC 50
51 CAATGGCAGC

Figure 12B (SEQ ID NO: 12)

MAAAIISVFSSGYSFYKQNLLLDSKPNILKPPCLLSPVVIMS PMRKKKKHGDPCICSVAGR
TRNLDIPQIEEEEEEENVEELIEQTDSDIVHIKKTLGGKQSKRPTGSIVAPVSCLGIIISMIC
PAVYFKFSRLMEGGDIPVAEMGITFATFVAAVGTEFLSAWVHKELWESLWYIHKSHHR
SRKGRFEFNDVFAIINALPAIALINYGFSNEGGLPGACFGVGLGTTVCGMAYIFLHNGLS
HRRFPVWLIANVPYFHKLAAAHQIHHSGKFQGVPFGLFLGPKELEEVRGGTEELERVISR
TTKRTQPST*

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Figure 13

At1 : -----	* 20 * 40 * 60 *	fkr fsvcyvve : 52 At2 : -----
At2 : -----	MAAGLSTIAVTLKPLNRSSFSANHPIstavffpslRFNGFRR-----rkiltvcfivve : 53	
Cal : -----	MAAEISISASSRAICLQRNPFPAKYFATAPPllffsplCNLDAILRSRRkprlaacvfk : 62	
Ca2 : TTGRYHYQLVWCQISFSSTSRTSYYRHSPFLGPKPTTPSVYpitpfspnlGSILRCRR---rpsitvcfivle : 71		
AdK1 : -----	AISVPSTS*PHKNLLHSKQDLNRP*llfspvvESMRKKKT-hraadicvae : 56	
AdK6 : -----	MAAAISVFSSGY*FYKNLLDSKPN*LKPP*llfspvvMSMRKKKK-hgdpcicvag : 59	
	FsP C V	
	S	
	80 * 100 * 120 * 140	
At1 : errqNSPIENDERPESTSSSTNAIDAEYLAL-----	rlaeeklenkkserstyliamissfgitsmavmavyy-fs : 122	
At2 : erkjSSPMDDDNKPESTTSSEILMTS-----	rllkkkaekkkserftyliavaavmssfgitsmavmavyy-fs : 120	
Cal : ddkLYTAQSGKQSDTEAIGDEIEVETNEEKSLAVnlaekfankksertfylvaavmsslgitsmavmavyy-fs : 136		
Ca2 : ddkfKTOFEAGEEDEIEMKIEEQISAT-----	laeklaakkserftyliavaavmssfgitsmavmavyy-fy : 137	
AdK1 : rjrnjdjorjEEEEEEENEEETEQTDSGII-----	rjkulgkksrrstwiajrvs.olgismmavmavyy-fs : 125	
AdK6 : rjrnjdjorjEEEEEEENVEETEQTDSDIV-----	rjkulgkkskrptwiajrvs.olgismmavmavyy-fs : 128	
	K S R T A S GI SM aVY Fs	
	160 * 180 * 200 * 220	
At1 : womeggeismlamfgtfalsvg-aavgefwarwahralwhaslwlmheshhkpregpfelndvfaivnagpai : 195		
At2 : wmkkggevsvlaemfgtfalsvgaaavvg-mefwarwahralwhdlslwlmheshhkpregalndvfaivnvpai : 194		
Cal : womeggeempfsemfctfallafg-aaigmeywarwahralwhaslwlmheshhkpregpfelndifaiinavpai : 209		
Ca2 : womeggevpfsemfctfallalsvg-aavgefwarwahkalwhaslwlmheshhkpregpfelndvfaivnvpai : 210		
AdK1 : wimecgrdipvaemdtfaafava-aaigbefmgwihkjlwhdlslwymhshhrfrg.fefndvfaiinalpai : 198		
AdK6 : wimeggrdipvaemdtfaafava-aavgefmgwihkjlwheslwymhshhrfrg.fefndvfaiinalpai : 201		
MegG EM TFA v Aa G Ef W H LWH SLW H SHH R G YE NDvFAI NA PAI		
	240 * 260 * 280 *	
At1 : gllsygffnkglvpglcfgaglgltvfigaymfvhgdlvhkrfpvggpiadvpvylrkvaahqjhhtckfnvgpy : 269		
At2 : gllyygflnkglvpglcfgaglgltmfgmaymfvhgdlvhkrfpvggpianvpvylrkvaahqjhhtckfkvgpy : 268		
Cal : affsfgfnhkglipgcfcfgaglgltvfigmaymfvhgdlvhkrfpvggpiakvpyfqrvaaahqjhhsckfdgvpy : 283		
Ca2 : alldygffhkglipgcfcfgaglgltvfigmaymfvhgdlvhkrfpvggvanvpvylrkvaahsjhhsckfnvgpy : 284		
AdK1 : alinygfsnigllpgcfgtglgtvfigmayfihnglshirfpvgianvpfyhkiaaahqjhhsckfqqgvp : 272		
AdK6 : alinygfsnigllpgcfvgqlgtvfigmayfihnglshirfpvwiianvpfyhkiaaahqjhhsckfqqgvp : 275		
1 yGF GL PG CFG GLG Tv GmAY P H GL H RFPVg ia VPY k AAAHQ HH KF GVP		
	300 * 320 *	
At1 : glflgpkeleevgg-nealdkeisriksykkasGSGSSSSS : 310		
At2 : glflgpqeveevgGkealekeisriklynkgSSTS----- : 305		
Cal : glflgpkeleevgv-iaealekevnrikslkrl----- : 315		
Ca2 : glflgpkeleevgg-lealekevnritryikgs----- : 316		
AdK1 : glflgpqeveevgGtaealekisrikqsst----- : 306		
AdK6 : glflgpkeleevgGtaealekisritkqsst----- : 309		
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SEQUENCE LISTING

<110> CUNNINGHAM, Francis X.

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<130> 8172-9022

<140> Unknown

<141> 1999-05-21

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<151> 1998-05-22

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ccttgatatt cctcaaattt aagaagagga agagaacgag gaagaactaa tagaacagac 240
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Pro Val Val Val Glu Ser Pro Met Arg Lys Lys Lys Thr His Arg Ala
35 40 45

Ala Cys Ile Cys Ser Val Ala Glu Arg Thr Arg Asn Leu Asp Ile Pro
50 55 60

Gln Ile Glu Glu Glu Glu Asn Glu Glu Glu Leu Ile Glu Gln Thr
65 70 75 80

Asp Ser Gly Ile Ile His Ile Lys Lys Thr Leu Gly Gly Lys Gln Ser
85 90 95

Arg Arg Ser Thr Gly Ser Ile Val Ala Pro Val Ser Cys Leu Gly Ile
100 105 110

Leu Ser Met Ile Gly Pro Ala Val Tyr Phe Lys Phe Ser Arg Leu Met
115 120 125

Glu Cys Gly Asp Ile Pro Val Ala Glu Met Gly Ile Thr Phe Ala Ala
130 135 140

Phe Val Ala Ala Ala Ile Gly Thr Glu Phe Leu Ser Gly Trp Val His
145 150 155 160

Lys Glu Leu Trp His Asp Ser Leu Trp Tyr Ile His Lys Ser His His
165 170 175

Arg Ser Arg Lys Gly Arg Phe Glu Phe Asn Asp Val Phe Ala Ile Ile
180 185 190

Asn Ala Leu Pro Ala Ile Ala Leu Ile Asn Tyr Gly Phe Ser Asn Glu
195 200 205

Gly Leu Leu Pro Gly Ala Cys Phe Gly Thr Gly Leu Gly Thr Thr Val
210 215 220

Cys Gly Met Ala Tyr Ile Phe Leu His Asn Gly Leu Ser His Arg Arg

225

230

235

240

Phe Pro Val Gly Leu Ile Ala Asn Val Pro Tyr Phe His Lys Leu Ala
245 250 255

Ala Ala His Gln Ile His His Ser Gly Lys Phe Gln Gly Val Pro Phe
260 265 270

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Ser Thr
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gagaaaagaaa aagaaacatg gtgatccatg tatctgctcc gttcagggaa gaacaaggaa 180
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20 25 30

Pro Val Val Ile Met Ser Pro Met Arg Lys Lys Lys His Gly Asp
35 40 45

Pro Cys Ile Cys Ser Val Ala Gly Arg Thr Arg Asn Leu Asp Ile Pro
50 55 60

Gln Ile Glu Glu Glu Glu Asn Val Glu Glu Leu Ile Glu Gln Thr
65 70 75 80

Asp Ser Asp Ile Val His Ile Lys Lys Thr Leu Gly Gly Lys Gln Ser
85 90 95

Lys Arg Pro Thr Gly Ser Ile Val Ala Pro Val Ser Cys Leu Gly Ile
100 105 110

Leu Ser Met Ile Gly Pro Ala Val Tyr Phe Lys Phe Ser Arg Leu Met
115 120 125

Glu Gly Gly Asp Ile Pro Val Ala Glu Met Gly Ile Thr Phe Ala Thr
130 135 140

Phe Val Ala Ala Ala Val Gly Thr Glu Phe Leu Ser Ala Trp Val His
145 150 155 160

Lys Glu Leu Trp His Glu Ser Leu Trp Tyr Ile His Lys Ser His His
165 170 175

Arg Ser Arg Lys Gly Arg Phe Glu Phe Asn Asp Val Phe Ala Ile Ile
180 185 190

Asn Ala Leu Pro Ala Ile Ala Leu Ile Asn Tyr Gly Phe Ser Asn Glu
195 200 205

Gly Leu Leu Pro Gly Ala Cys Phe Gly Val Gly Leu Gly Thr Thr Val
210 215 220

Cys Gly Met Ala Tyr Ile Phe Leu His Asn Gly Leu Ser His Arg Arg
225 230 235 240

Phe Pro Val Trp Leu Ile Ala Asn Val Pro Tyr Phe His Lys Leu Ala

245

250

255

Ala Ala His Gln Ile His His Ser Gly Lys Phe Gln Gly Val Pro Phe
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Ser Thr
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35 40 45

Met Arg Lys Lys Lys Thr His Arg Ala Ala Cys Ile Cys Ser Val Ala
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Glu Arg Thr Arg Asn Leu Asp Ile Pro Gln Ile Glu Glu Glu Glu
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85 90 95

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Val Tyr Phe Lys Phe Ser Arg Leu Met Glu Cys Gly Asp Ile Pro Val
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Ala Glu Met Gly Ile Thr Phe Ala Ala Phe Val Ala Ala Ala Ile Gly
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Thr Glu Phe Leu Ser Gly Trp Val His Lys Glu Leu Trp His Asp Ser
165 170 175

Leu Trp Tyr Ile His Lys Ser His His Arg Ser Arg Lys Gly Arg Phe
180 185 190

Glu Phe Asn Asp Val Phe Ala Ile Ile Asn Ala Leu Pro Ala Ile Ala
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Leu Ile Asn Tyr Gly Phe Ser Asn Glu Gly Leu Leu Pro Gly Ala Cys
210 215 220

Phe Gly Thr Gly Leu Gly Thr Thr Val Cys Gly Met Ala Tyr Ile Phe
225 230 235 240

Leu His Asn Gly Leu Ser His Arg Arg Phe Pro Val Gly Leu Ile Ala
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Asn Val Pro Tyr Phe His Lys Leu Ala Ala Ala His Gln Ile His His
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Ser Gly Lys Phe Gln Gly Val Pro Phe Gly Leu Phe Leu Gly Pro Gln
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Gly Arg Thr Arg Asn Leu Asp Ile Pro Gln Ile Glu Glu Glu Glu
65 70 75 80

Asn Val Glu Glu Leu Ile Glu Gln Thr Asp Ser Asp Ile Val His Ile
85 90 95

Lys Lys Thr Leu Gly Gly Lys Gln Ser Lys Arg Pro Thr Gly Ser Ile
100 105 110

Val Ala Pro Val Ser Cys Leu Gly Ile Leu Ser Met Ile Gly Pro Ala
115 120 125

Val Tyr Phe Lys Phe Ser Arg Leu Met Glu Gly Gly Asp Ile Pro Val
130 135 140

Ala Glu Met Gly Ile Thr Phe Ala Thr Phe Val Ala Ala Ala Val Gly
145 150 155 160

Thr Glu Phe Leu Ser Ala Trp Val His Lys Glu Leu Trp His Glu Ser
165 170 175

Leu Trp Tyr Ile His Lys Ser His His Arg Ser Arg Lys Gly Arg Phe
180 185 190

Glu Phe Asn Asp Val Phe Ala Ile Ile Asn Ala Leu Pro Ala Ile Ala
195 200 205

Leu Ile Asn Tyr Gly Phe Ser Asn Glu Gly Leu Leu Pro Gly Ala Cys
210 215 220

Phe Gly Val Gly Leu Gly Thr Thr Val Cys Gly Met Ala Tyr Ile Phe
225 230 235 240

Leu His Asn Gly Leu Ser His Arg Arg Phe Pro Val Trp Leu Ile Ala
245 250 255

Asn Val Pro Tyr Phe His Lys Leu Ala Ala Ala His Gln Ile His His

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265

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Ser Arg Thr Thr Lys Arg Thr Gln Pro Ser Thr
305 310 315

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Arg Pro Glu Ser Thr Ser Ser Thr Asn Ala Ile Asp Ala Glu Tyr Leu
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Thr Tyr Leu Ile Ala Ala Met Leu Ser Ser Phe Gly Ile Thr Ser Met
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Glu Ile Ser Met Leu Glu Met Phe Gly Thr Phe Ala Leu Ser Val Gly
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Trp His Ala Ser Leu Trp Asn Met His Glu Ser His His Lys Pro Arg
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Pro Ala Ile Gly Leu Leu Ser Tyr Gly Phe Phe Asn Lys Gly Leu Val
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Pro Gly Leu Cys Phe Gly Ala Gly Leu Gly Ile Thr Val Phe Gly Ile
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Ala Tyr Met Phe Val His Asp Gly Leu Val His Lys Arg Phe Pro Val
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Gly Pro Ile Ala Asp Val Pro Tyr Leu Arg Lys Val Ala Ala His
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Gln Leu His His Thr Asp Lys Phe Asn Gly Val Pro Tyr Gly Leu Phe
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Glu Gln Thr Asp Ser Asp Ile Val His Ile Lys Lys Thr Leu Gly Gly
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His Arg Arg Phe Pro Val Trp Leu Ile Ala Asn Val Pro Tyr Phe His
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Trp His Ala Ser Leu Trp Asn Met His Glu Ser His His Lys Pro Arg
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Glu Gly Pro Phe Glu Ieu Asn Asp Val Phe Ala Ile Val Asn Ala Gly
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Pro Ala Ile Gly Leu Leu Ser Tyr Gly Phe Phe Asn Lys Gly Leu Val
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Pro Gly Leu Cys Phe Gly Ala Gly Leu Gly Ile Thr Val Phe Gly Ile
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Ala Tyr Met Phe Val His Asp Gly Leu Val His Lys Arg Phe Pro Val
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Gly Pro Ile Ala Asp Val Pro Tyr Leu Arg Lys Val Ala Ala His
245 250 255

Gln Leu His His Thr Asp Lys Phe Asn Gly Val Pro Tyr Gly Leu Phe
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Leu Gly Pro Lys Glu Leu Glu Val Gly Gly Asn Glu Glu Leu Asp
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Ser Val Leu Glu Met Phe Gly Thr Phe Ala Leu Ser Val Gly Ala Ala
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Val Val Gly Met Glu Phe Trp Ala Arg Trp Ala His Arg Ala Leu Trp
145 150 155 160

His Asp Ser Leu Trp Asn Met His Glu Ser His His Lys Pro Arg Glu
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Gly Ala Phe Glu Leu Asn Asp Val Phe Ala Ile Thr Asn Ala Val Pro

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Gly Leu Cys Phe Gly Ala Gly Leu Gly Ile Thr Met Phe Gly Met Ala
210 215 220

Tyr Met Phe Val His Asp Gly Leu Val His Lys Arg Phe Pro Val Gly
225 230 235 240

Pro Ile Ala Asn Val Pro Tyr Leu Arg Lys Val Ala Ala Ala His Gln
245 250 255

Leu His His Thr Asp Lys Phe Lys Gly Val Pro Tyr Gly Leu Phe Leu
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Gly Asp Glu Ile Glu Val Glu Thr Asn Glu Glu Lys Ser Leu Ala Val
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Arg Leu Ala Glu Lys Phe Ala Arg Lys Lys Ser Glu Arg Phe Thr Tyr
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Leu Val Ala Ala Val Met Ser Ser Leu Gly Ile Thr Ser Met Ala Val
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Ile Ser Val Tyr Tyr Arg Phe Ser Trp Gln Met Glu Gly Gly Glu Met
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Ile Gly Met Glu Tyr Trp Ala Arg Trp Ala His Arg Ala Leu Trp His
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Ala Ser Leu Trp His Met His Glu Ser His His Arg Pro Arg Glu Gly
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Pro Phe Glu Leu Asn Asp Ile Phe Ala Ile Ile Asn Ala Val Pro Ala
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Ile Ala Phe Phe Ser Phe Gly Phe Asn His Lys Gly Leu Ile Pro Gly
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Ile Cys Phe Gly Ala Gly Leu Gly Ile Thr Val Phe Gly Met Ala Tyr
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Met Phe Val His Asp Gly Leu Val His Lys Arg Phe Pro Val Gly Pro
245 250 255

Ile Ala Lys Val Pro Tyr Phe Gln Arg Val Ala Ala Ala His Gln Leu
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His His Ser Asp Lys Phe Asp Gly Val Pro Tyr Gly Leu Phe Leu Gly
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Thr Val Cys Phe Val Leu Glu Asp Asp Lys Phe Lys Thr Gln Phe Glu
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Ala Gly Glu Glu Asp Ile Glu Met Lys Ile Glu Glu Gln Ile Ser Ala
85 90 95

Thr Arg Leu Ala Glu Lys Leu Ala Arg Lys Lys Ser Glu Arg Phe Thr
100 105 110

Tyr Leu Val Ala Ala Val Met Ser Ser Phe Gly Ile Thr Ser Met Ala
115 120 125

Val Met Ala Val Tyr Tyr Arg Phe Tyr Trp Gln Met Glu Gly Glu
130 135 140

Val Pro Phe Ser Glu Met Phe Gly Thr Phe Ala Leu Ser Val Gly Ala
145 150 155 160

Ala Val Gly Met Glu Phe Trp Ala Arg Trp Ala His Lys Ala Leu Trp
165 170 175

His Ala Ser Leu Trp His Met His Glu Ser His His Lys Pro Arg Glu
180 185 190

Gly Pro Phe Glu Leu Asn Asp Val Phe Ala Ile Ile Asn Ala Val Pro
195 200 205

Ala Ile Ala Leu Leu Asp Tyr Gly Phe Phe His Lys Gly Leu Ile Pro
210 215 220

Gly Leu Cys Phe Gly Ala Gly Leu Gly Ile Thr Val Phe Gly Met Ala
225 230 235 240

Tyr Met Phe Val His Asp Gly Leu Val His Lys Arg Phe Pro Val Gly
245 250 255

Pro Val Ala Asn Val Pro Tyr Leu Arg Lys Val Ala Ala Ala His Ser
260 265 270

Leu His His Ser Glu Lys Phe Asn Gly Val Pro Tyr Gly Leu Phe Leu
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32

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US99/10455

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :C12P 23/00, 7/26; C12N 9/02, 1/20, 15/00; C07H 21/04; C07K 14/00
US CL :435/67, 148, 189, 252.3, 252.33, 320.1; 536/23.2, 23.6; 530/350

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/67, 148, 189, 252.3, 252.33, 320.1; 536/23.2, 23.6; 530/350

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	US 5,453,565 A (MAWSON) 26 September 1995, see abstract and claims.	1-20
A,E	US 5,910,433 A (KAJIWARA et al.) 08 June 1999, see the entire patent.	1-20
Y,P	US 5,811,273 A (MISAWA et al.) 22 September 1998, See abstract, column 30 - lines 48-58 and claims.	1-20

Further documents are listed in the continuation of Box C.

See patent family annex.

Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
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O document referring to an oral disclosure, use, exhibition or other means		
P document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

13 AUGUST 1999

Date of mailing of the international search report

29 OCT 1999

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231

Authorized Officer

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Faximile No. (703) 305-3230

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US99/10455

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, STN Files: Medline, Caplus, Biosis, Agricola, Embase & Scisearch. Search terms used : beta carotene and ketolase, ketocarotenoid, *Adonis aestuialis*, carotenoid biosynthesis, gene? or dna or rna or nucleic acid? in various permutations and combinations.

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